



Research Paper

The low-temperature stress tolerance of the grape varieties of ecological and geographical origin

Accepted 15th November, 2018

ABSTRACT

The cold-tolerance of 'Kristal' (Euro-Amur-American origin), 'Dostoyiniy' and 'Krasnostop AZOS' (interspecific hybrids of Euro-American origin) grapevine varieties in the conditions of the Russian South winter is characterized by the second (true dormancy) and third (induced dormancy) winter-resistance components. Here, a complex approach is used to estimate the plants adaptability to abiotic stress factors. We used gravimetric method for humidity assessment and water content; spectral analysis for detection of the protein level and pigments; capillary electrophoresis method to analyze the level of carbohydrates, Krebs cycle organic acids, phenolcarbonic, ascorbic acids, and amino acids. The obtained results allow to suppose the various mechanisms of adaptation of the studied varieties to the winter period stressors. The water content of the shoots, the level of protein, aminoacids, proline, sugars and the sum of phenolcarbonic and ascorbic acids are the most informative indicators of grape plants frost-resistance in the climate conditions of Anapa - Taman region.

Nataliya Ivanovna NENKO*, Irina Anatolievna ILYINA, Galina Konstantinovna KISELEVA and Elena Karlenovna YABLONSKAY

North Caucasian Regional Research
Institute of Horticulture and Viticulture.
39, 40 let Pobedy Street, Krasnodar
350901, Russia.

*Corresponding author. E-mail:
nenko.ncrrihv@bk.ru. Tel:
+79231594735.

Key words: grape varieties, frost tolerance, interspecific hybrids, abiotic stress, adaptations, metabolic evaluation.

INTRODUCTION

The understanding of the mechanisms of grape plants cold-resistance to abiotic and biotic factors of the environment have a relevance for the agricultural industry and biotechnology of highly productive grape varieties (Ollat et al., 2017; Nenko et al., 2017; Trejo-Martinez et al., 2009; Arun-Chinnappa et al., 2017). The natural and climatic conditions of the South of Russia are favourable to ensure that the high yields of grape are able to withstand the competitions on a worldwide basis. At the same time, producing stable high yields is restricted by the influence of such unfavourable environmental factors, as winter frosts, especially after the long warm weather in summer period. That is why only the varieties, bringing together high productivity, good quality and adaptability to conditions of this region, may be successfully cultivated here on a

sufficiently broad scale. It dictates the need for improved range of varieties through the cultivation of grape varieties, better adapted to the weather conditions of the habitats (Ferrandino and Lovisolo, 2014; Sha Valli et al., 2014). A key issue of adaptation is to study the realization of the potential, genetically determined organism abilities in response to the action of unfavourable environmental factors (Nenko et al., 2017; Yegorov et al., 2017).

The complex estimate of the grape plants based on the physiological and biochemical indicators, which may be used as the indirect diagnostic techniques, permits to obtain more reliable characteristic of the genotypes resistance to the unfavourable weather and climatic conditions (Xian et al., 2017; Arun-Chinnappa et al., 2017). In this regard, the study of physiological and biochemical

mechanisms of adaptation of the grape plants, providing the orderliness and regulation of the physiological processes, is especially topical (Nenko et al., 2015; Beheshti Rooy et al., 2017; Zhang et al., 2012., Tsvica et al., 2007, Trejo-Martinez et al., 2009)).

The objective of this work was to study the physiological and biochemical mechanisms of low-temperature resistance of grape varieties of different ecological and geographical origin during winter period and identify varieties of low-temperature resistance during winter period on the basis of metabolic evaluation of genotype expression in hydrothermal conditions of the Russian South.

MATERIALS AND METHODS

The minimum temperature on the territory of the Anapa-Taman zone of the South of Russia (Figure 1) in the winter 2014-2015 was -7°C in December 2014, -19°C in January, and -11°C in February, with precipitation quantity of - 81, 73 and 15 mm, respectively. The minimum air temperature in the winter period of 2015-2016 reached -9°C in December, -13°C in January and -5°C in February, with precipitation quantity to 28, 118 and 47 mm, respectively.

The study was done during the period from December to February of 2014 - 2016 on the territory of Anapa. During this period, there was an increase in the minimum air temperature in January and February by 6°C and in precipitation quantity- by 45 and 32 mm, respectively. During March of the same period, the precipitation quantity was 13 mm, which is an evidence of gradual warming conditions.

The research was conducted on the basis of ampelographic collection of the Federal state budgetary scientific institution "Anapa Zonal Pilot Station of Viniculture and Wine-making", located in Anapa city, block of the grape vine industrial varieties on southern calcareous chernozem. The research was carried out using the equipment of the Centre of collective usage "Tool and Analytical" and physiology and biochemistry laboratory of the Federal state budgetary scientific institution "North-Caucasian federal scientific centre of horticulture, viniculture and winemaking" (Nenko et al., 2017; Yegorov et al., 2017).

The plants were of the same implantation year (1995), parent stock was Kober 5BB. Pruning for shape is a bilateral high-standard spiral cordon of Anapa Zonal Pilot Station. Cultivation of grape plants was performed in bare fallow with landing procedure 3×2.5 m. The objects of research were the grape vine varieties of different ecological and geographic origin and maturation periods, survived after extreme temperatures of the year 2012 winter (Yegorov et al., 2017).

The 'Kristall' is a variety, the plant of early maturation period, was used as a control (interspecific hybrid of Euro-Amur-American origin). 'Dostoiniy' and 'Krasnostop' are medium maturation period varieties of Anapa Zonal Pilot Station (interspecific hybrids of Euro-Amur-American origin) (Yegorov et al., 2017).

The 'Kristall' is a variety of Hungarian breeding, selected for the industrial purposes. The 'Kristall' parental pair is 'Amurskiy' X Csalloci Lojos and Villar Blan Hungarian variety. This 'Kristall' is high-productive variety characterized by very early maturation period and frost-resistance (up to the -27°C). The 'Kristall' grapes have yellowish-green color and harmonious taste. The bunch of 'Kristall' variety have medium density, weighing 180-200 g. The sugar and acidity content ranges from 17 to 18%, and from 6 to 7 g/L, respectively (Yegorov et al., 2017).

'Dostoiniy' variety was selected by the staff of Anapa Zonal Pilot Station for the industrial purposes. The parental pair is phylloxera-resistant Jemete X Muscat Hamburg. The 'Dostoiniy' variety has medium maturation period. Phylloxera-resistant may be grown on its own roots, but it is highly susceptible to mildew and oidium. Bunches are of conical form with medium density and medium size. The taste is ordinary; its berries are used to prepare the dry wines. (Yegorov et al., 2017).

'Krasnostop AZOS' variety was also selected by Anapa Zonal Pilot Station breeders for industrial purposes and wine production. Its parental pair is phylloxera-resistant Jemete X Krasnostop Anapsky. It has medium early maturation period. On an average the acidity of grapes juice is 8 g/L, the sugar content is 24%. The form of 'Krasnostop AZOS' grapevines is cylindrical. The bunches are of medium density and small, about 120-130 g in weight (Yegorov et al., 2017).

To estimate the adaptation resistance of the grape vine plants to abiotic stresses, humidity, free and bound water content, we used the weight method (Kushnirenko and Pecherskaya, 1991). To determine the content of free water, the specimens of shoots weighing 1 g each were dried in three replications for 24 h on open air, after which they were weighed. To determine the content of free water, the weight difference was referred to the weight of total water content. The 1 g weighed quantities were dried in three replications in thermostat until the weight becomes constant at 105°C to determine the total content of water in the shoots. The water content of shoots was determined as a ratio between difference of the raw weight of shoots and its dry mass to the weight of raw biomass. The content of bound water within the specimens was found as the difference between the shoots water content and free water content and expressed as a percentage. The protein and pigments content was determined using a spectral method (Yermakov et al., 1972).

To determine the protein content in the shoots, the 1 g



Figure 1: The location of the territory of the Anapa-Taman zone of the South of Russia.

specimens were sampled in three replications. The specimens were ground in cryogenic nitrogen followed by protein extraction. For extraction of protein, we used the buffer solution, containing in 0.48 mg 0.1 M Tris, 0.1 g ascorbic acid, 0.08 g ethylenediaminetetraacetate, 0.18 g sodium diethyldithiocarbamate, 0.145 g sodium chloride, 2.0 g polyethyleneglycol and distilled water in a total of 100 ml volume. Samples were homogenized and stored in a buffer for at least 2 h. After that samples were centrifuged in the cold (+4°C, centrifuge 5418 R eppendorf), the centrifugate was poured off into the separate test glasses, and the precipitate was repeatedly topped up with the buffer. The extraction procedure was repeated three times, the precipitate of each replicate was combines in 100 ml flasks with the addition of a buffer to the label. The obtained solution was poured into the 10 mm thick quartz cells. The optical density of solutions was determined by UNICO 2800 spectrophotometer at 280 nm wavelength. Finally, the protein content of specimen was determined by calibration curve (Yermakov et al., 1972).

To estimate the content of pigments (anthocyanins, chalcones) in the shoots, 1 g specimens were ground in cryogenic nitrogen, flooded with 10 ml of 0.1 n hydrochloric acid solution, infused in the cold for 1 h and centrifugated. The extraction was performed three times. The supernatant was put into 100 ml flasks, completed with 0.1 n hydrochloric acid solution to the mark. The anthocyanins and chalcones content were determined on LEKI SS1207 spectrophotometer at 490 and 364 nm wavelength, respectively (Yermakov et al., 1972). The starch content was determined using polarimetric method on universal polarimeter (Yermakov et al., 1972).

The level of carbohydrates (sucrose), Krebs cycle organic acids (malic, citric, succinic), phenolcarbonic acids (chlorogenic, caffeic), ascorbic and proline amino acid was done by capillary electrophoresis. As a result, the 1 g

specimen was ground in three replications, each one in liquid nitrogen. It was then put into fluoroplastic container of “Minotaur” microwave mineralizer, 25 ml of 10% alcohol (ethanol 98 %) aqueous rectified alcohol were added. The container was placed into mineralizer magnetron and processed for 10 min in “pressureless decomposition” mode, with the application of 10% mineralizer magnetron capacity. After that, the container was removed from microwave mineralizer, cooled under natural conditions for 5 min and the obtained liquid was quantitatively taken into the 25 ml graduated flasks by adding the initial alcohol-water mixture. Further, the content of these components were determined quantitatively by Kapel 105 P tool, equipped with UV photometric detector with the addition of the initial alcohol-water mixture (Yakuba et al., 2015a, b, c, d; The methodological and analytical support of research in horticulture, 2010).

Furthermore, the content of these components were determined quantitatively by Kapel 105 P tool, equipped with UV photometric detector, operating at 254 nm wavelength, quartz capillary at 0.5m length at least to the detector, 50-100 µm inner diameter, positive polarity high-voltage source of variable voltage ranging from 1 to 25 kV and personal computer with the relevant software for the collection and processing of data. The target values of the analyzed substances content were determined with the parameters as follows: positive voltage on capillary – 16 kV, time of analysis - 15 min for ascorbic, chlorogenic and caffeic acids. The individual connections were identified using the standard addition method.

For organic acids (malic, citric, succinic), the operating parameters were laid down as follows: the wavelength of spectrophotometric detector – 270 nm, negative voltage – 25 kV, pneumatic dosage of sample at 30 mbar for 5 s at 0 kV voltage, time of analysis – 10 min. The terms of the samples electrophoregram registration met those of

calibration solutions electrophoregram registration. The individual connections were identified by the standard addition method. The target values of the analyzed substances content were determined with the parameters as follows: positive voltage on capillary – 10 kV, time of analysis - 40 min.

For the determination of amino acid, to 0.05 ml of extract was added 0.1 ml of a 10% aqueous solution of sodium carbonate and 0.3 ml of a solution phenylisothiocyanate in isopropyl alcohol and left for 35 min to undergo reaction. The resulting solution was dried, dissolved in 0.5 ml of distilled water, centrifuged for 5 min and transferred to the device under a pressure of 30 millibars for 5 s. The individual connections were identified using the standard addition method.

The analysis employed aqueous leading electrolyte, containing 0.33% of boric acid, 0.05% sodium tetraborate and 0.5% isopropylalcohol with the positive polarity of voltage and the length of detection wave – 254 nm.

The preparation of samples for morphological studies involved commonly used botanic micro methods (The methodological and analytical support of research in horticulture, 2010).

The measurement data were processed with the use of conventional methods of variation statistics (Dospikhov, 1979; Welham et al., 2014). The preparation of samples for morphological studies involved commonly used botanic microengineering methods (Yakuba et al., 2015a). The plants resistance to low temperatures of winter period was studied under the natural conditions as well as conditions of induced stress, including forced dehydration and low temperature (-25°C). The experimental data were analyzed by the commonly used methods of variation statistics (Yakuba et al., 2015b). The studies were carried out with the use of filter photometer; MBI-3, MBI-10 and Olympus microscopes; UNICO 2800 and LEKI SS1207 spectrophotometers; Kapel 105 P capillary electrophoresis tool; JW-1-3000 Acom balance and analytic balance; LE-402, Type-310, TsLN-16 centrifuges; Eppendorf 5418R LOIP LB-163 (TE-6/24-BK) water bath; SESH-1 drying cupboard; Gronland refrigerating cabinet.

RESULTS

All the analyzed varieties showed lower water content of shoots and their content of free water in December of 2015, as compared with December of 2014. For vegetative season of 2015 as compared with 2014, the grape plants accumulated the macronutrients stored in the shoots. The level of water increased by 13.72% for 'Kristall' variety and by 6.1- 9.0% for 'Dostoyiniy' and 'Krasnostop' AZOS varieties, respectively (Figure 2)

In December of 2015 as compared with 2014, free-water contents of 'Kristall' and 'Krasnostop AZOS' varieties, along

with the lesser water content of shoots, lowered (29.4 and 7.2%, respectively) and slightly increased in 'Dostoyiniy' variety (1.2%), prompting the suggestions that its metabolic behavior is more active.

During the period of 2012 – 2015, the free-water contents of shoots from 'Kristall' variety in December correlates with the maximum air temperature ($r=0.94$) and precipitation depth ($r=0.92$), and in 'Dostoyiniy' and 'Krasnostop AZOS' varieties – both to the maximum and minimum air temperature ($r=0.65 - 0.73$) and precipitation depth ($r=0.66 - 0.68$) (Figure 3).

In the December of 2015 as compared with 2014, the contents of chalcones, protecting the cell membranes from the breakage (Nenko et al., 2017), in the vine grape varieties under study grew by 63.7 – 159.9% (Figure 4). Consequently, the increase in chalcones contents of the grape vine is a common pattern for all the studied varieties.

The determination of starch in vine, characterizing the frost resistance in the true dormancy period, showed its lower contents in 'Kristall' variety (1.67 mg/g) in the December of 2015 and higher one in 'Dostoyiniy' variety (4.98 mg/g), and 'Krasnostop AZOS' variety (4.2 mg/g) holds intermediate position between these. The ratios of saccharose contents to that of starch in 'Kristall' variety in December of 2015 was 3.8, 'Dostoyiniy' – 0.96 and 'Krasnostop AZOS' – 0.55, characterizing 'Kristall' variety as a more frost-proof one. In the December of 2015, the bound water contents of the grape varieties' shoots were mostly influenced by sucrose contents ($r=0.7$). In the December of 2014, the desiccation resistance of the grape plants of the studied varieties was mostly influenced by proline contents (in 2013 proline and sucrose), characterizing the impact of year conditions on specificity of protective response to the stressors in the state of true dormancy (Figure 5).

The estimation of mechanism of the vine's desiccation resistance showed proline to have greater effect and sucrose to have lesser one on the vine's bound water contents ($r=0.5$) in 'Kristall' variety ($r=0.9$), in 'Krasnostop AZOS' variety – both proline and sucrose ($r=0.8$), and in 'Dostoyiniy' variety – proline and sucrose to have a slight impact ($r=0.3$) (Figure 6).

It brings us to the assumption of the different mechanisms of adaptation to the varied environment in the plants of varieties under study in the state of true dormancy. For the 2012-2015 period in December, the proline contents in the vine of 'Kristall' variety was higher (12.6 – 49.8 mg/kg) than in 'Krasnostop AZOS' variety (7.9 – 34.5 mg/kg) and 'Dostoyiniy' (7.6 – 30.4 mg/kg). In the December of 2015, 'Kristall' and 'Dostoyiniy' varieties are characterized by the higher contents of ascorbic acid (20.6 and 12.3 mg/kg, respectively) and the phenolcarbonic acids totality (51.2 and 62.5 mg/kg, respectively), protecting the cell membranes from destruction, and 'Krasnostop AZOS' variety – by the lower one (9.3 and 25.1 mg/kg, respectively) (Figure 7) (Nenko et al., 2017; Xian et al., 2017;

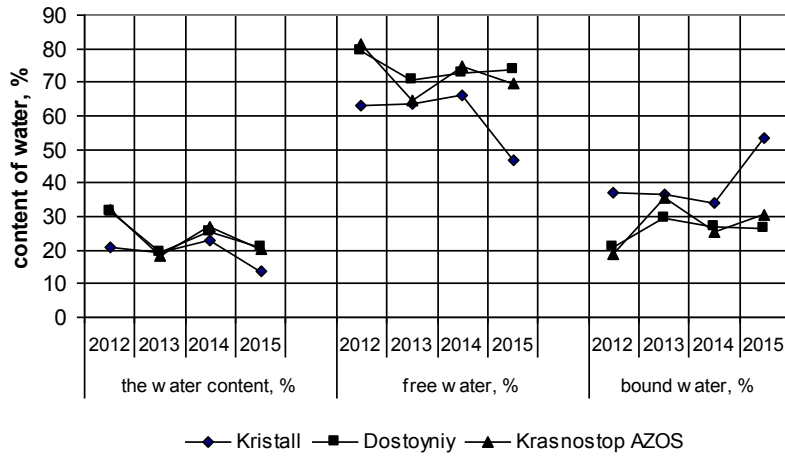


Figure 2: Moisture status of the grape shoots in December of 2012 - 2015.

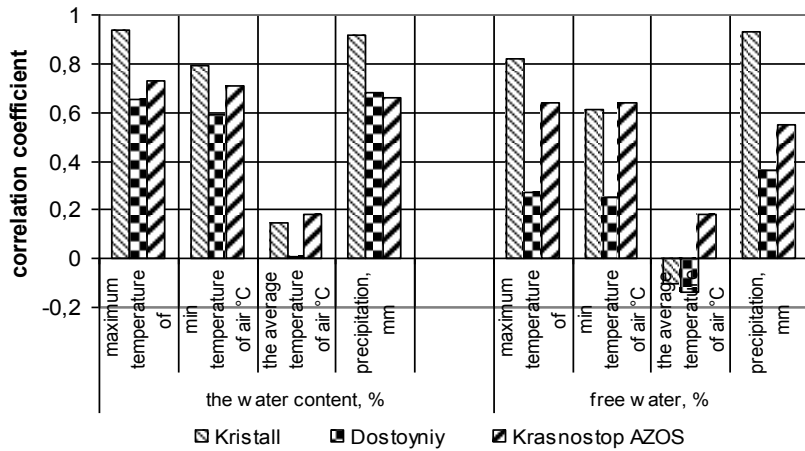


Figure 3: The dependence of the grape shoots' moisture status in 2012-2015 December hydrothermal conditions in Anapa -Taman zone.

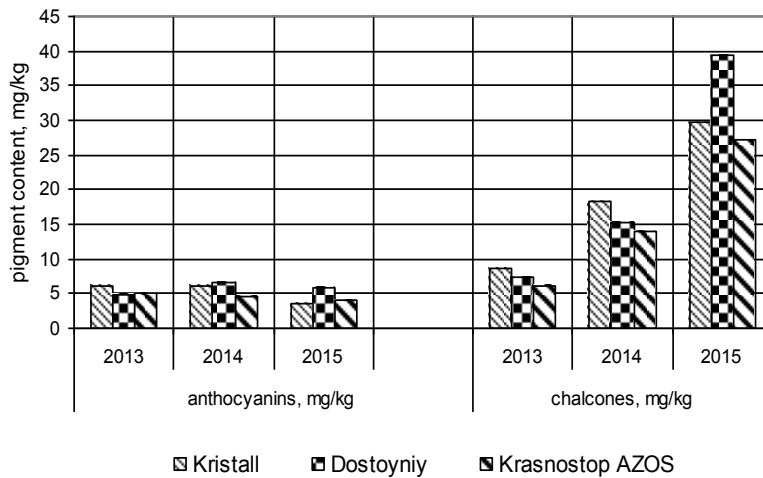


Figure 4: The pigments' contents of the grape vine in December of 2012-2015.

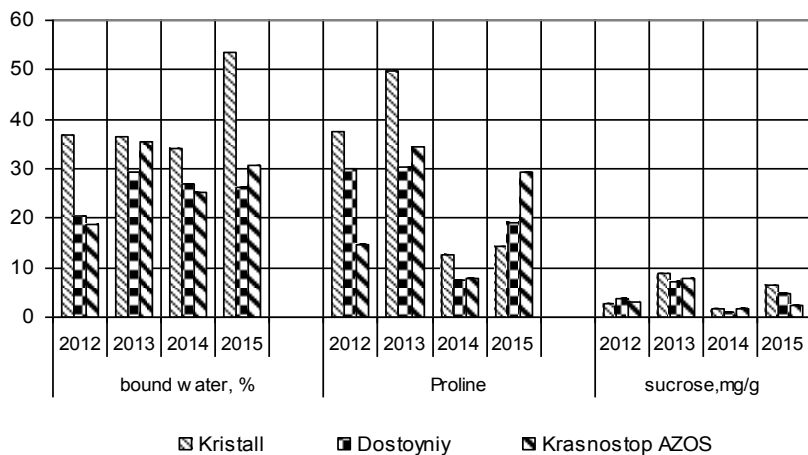


Figure 5: Water-retaining capacity of the grape vine in December of 2012-2015.

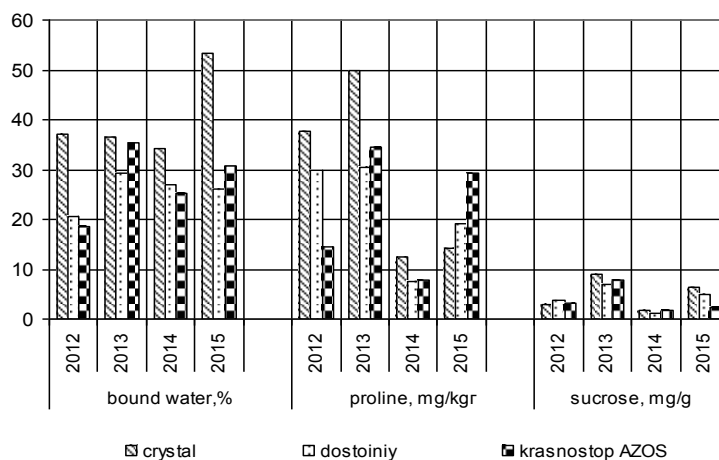


Figure 6: Biochemical characteristic of the grape shoots' water-retaining capacity in December of 2012-2015.

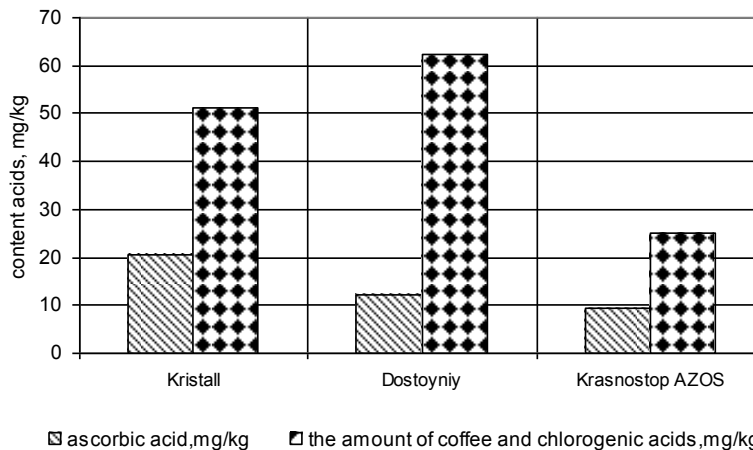


Figure 7: The ascorbic and phenolcarmonic acids contents of the grape shoots in December of 2015.

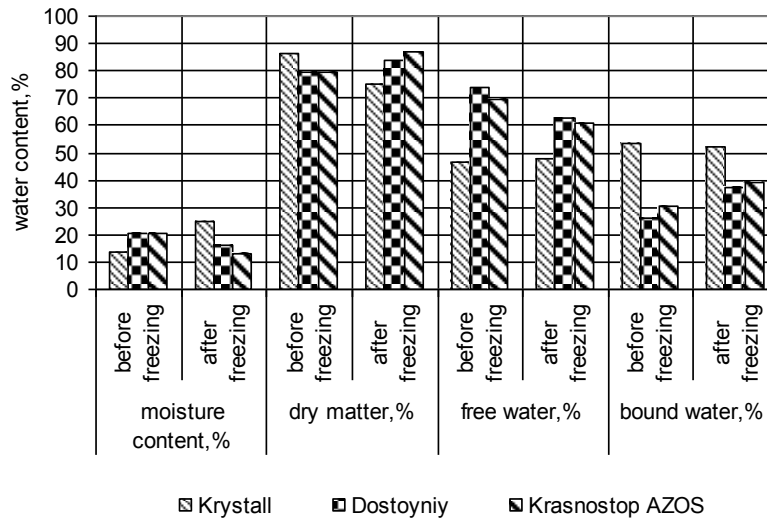


Figure 8: The impact of the low-temperature stress in model experiment on the water status of grape shoots, December of 2015.

Davey et al., 2000; Swanson and El-Shishiny, 1958; Wample and Bary, 1992). The histochemical studies established the starch contents to be 5.0 points for the fine-cellular core zone of 'Kristall' and 'Dostoyniy' varieties, and 4.7 points for 'Krasnostop AZOS' variety, permitting to characterize these as the highly frost-proof ones and frost-proof one, respectively. The frost-proof grape varieties are characterized by the fast hydrolysis of starch already from the beginning of winter. The hydrolysis of starch is delayed in the insufficiently frost-proof varieties.

It was found that the content of starch in the vine of 'Dostoyniy' and 'Kristall' varieties was 5.0 and 4.9 points, respectively in December as compared with November of 2015. The starch contents in the shoot fine-cellular core zone did not change for 'Krasnostop AZOS' variety. Thus, 'Dostoyniy' and 'Kristall' varieties showed their high winter hardiness, judging from the rates of starch consumption caused by its hydrolysis. 'Krasnostop AZOS' variety showed itself as frost-proof by the rate of starch consumption. The anatomo-morphological study of buds showed them to be in the state of winter dormancy. All the grape varieties had embryonic inflorescences, responsible for the next year yield, embedded in the winter buds (eyes) (Takahama and Oniki, 2000).

The artificial freezing of the grape shoots in December of 2015 at temperature of minus 25°C increased water content of the 'Kristall' variety shoots by 173%, and in 'Dostoyniy' and 'Krasnostop AZOS' varieties it dropped by 29.9 and 60.3%, respectively (Figure 8).

In such a case, the free water contents of these varieties lowered by 18.2 and 13.8%, respectively. The ratio of bound water to free water content after freezing was higher by 66.7% for 'Dostoyniy' variety and by 45.5% for

'Krasnostop AZOS' variety, respectively. The same ratio changed insignificantly for 'Kristall' variety. Freezing increased the protein contents of a 'Kristall' variety vine by 19.9%, it lowered in 'Dostoyniy' variety by 22.1% and 'Krasnostop AZOS' variety – by 28.6%, and starch contents – in 'Kristall' variety – by 36.5%, and as for 'Dostoyniy' and 'Krasnostop AZOS' varieties – by 175 and 96.3%, respectively (Figure 9). This goes to prove active hydrolytic behavior that agrees with the higher proline contents of 'Krasnostop AZOS' variety by 197.6%.

The contents of the Krebs cycle organic acids in 'Dostoyniy' variety was 52.2% lower that reflects the decrease in respiration intensity. Thus, we may assume, that in the December of 2015 the grape plants of this variety did not enter the state of deep dormancy. The elevated level of organic acids was detected in 'Kristall' and 'Krasnostop AZOS' varieties, 50.0% and 17.8% higher, respectively, displaying the protective response of plants through the activation of breathing.

The impact of the low-temperature on the contents of phenolcarboxylic acids in the grape vine of the studied varieties was also assessed. When freezing the shoots, the increase in the contents of the phenolcarboxylic acids (chlorogenic, caffeic) sum is reported for 'Kristall' variety by 135.4%, for 'Dostoyniy' – by 45.6% and 'Krasnostop AZOS' variety – 6.4 times more. It characterizes the different mechanisms of the extremely low temperature resistance of the grape varieties varying in ecological and geographical origin in the period of true dormancy.

The histochemical analysis demonstrated that after freezing of the shoots, the starch contents of the fine-cellular core zone of grape vine in 'Dostoyniy', 'Krasnostop AZOS' and 'Kristall' varieties, observed as the highly frost-proof

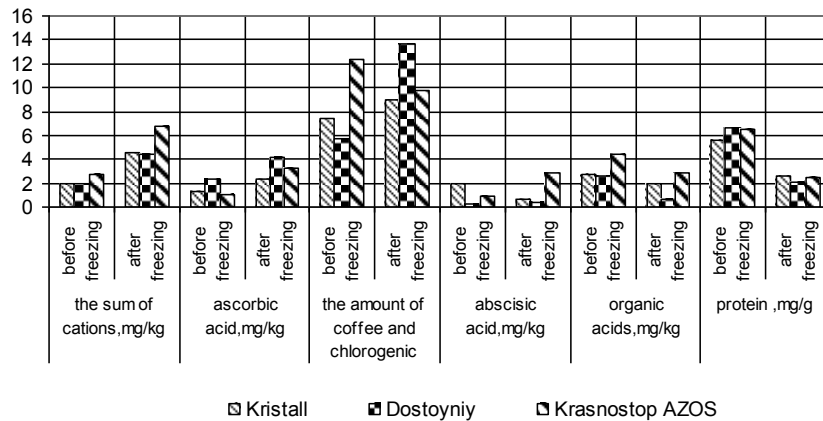


Figure 9: Biochemical characteristic of the grape vine frost resistance, December 2015.

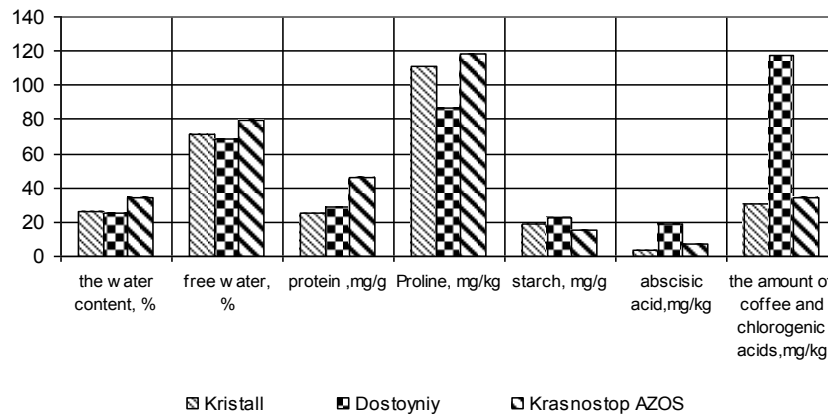


Figure 10: Biochemical characteristic of grape vine on the basis of the third winter-resistance component, February of 2016.

ones, did not change and continued to be 4.7 - 5.0 points. Thus, the influence of the low temperatures on the shoots of the studied grape varieties caused the largest changes in the vine cortex and did not damage its core.

Under the natural conditions of Anapa and Taman regions, the air temperature in January of 2015 dropped down to minus 19°C, which affected adversely the plants of grape in the state of induced dormancy. In late February of 2016, the water content of the shoots grew, as compared with December 2015 in 'Kristall' variety by 12.6%, 'Dostoynyi' – 22.2%, and 'Krasnostop AZOS' – 68.8%. The free water contents in 'Kristall' variety increased by 53.9%, in 'Dostoynyi' variety - 7.7%, and in 'Krasnostop AZOS' variety – by 14.8%, which are due to activation of metabolic processes.

In February of 2015, the contents of proline in vine increased by 4.1 – 7.7 times, and the level of starch diminished by 63.65 and 49.05%. This agrees with the

activation of the hydrolytic processes (Figure 10).

DISCUSSION

The study was conducted in the winters of 2012-2016. During these periods, the grape plants did not enter the state of deep dormancy and lacked the tolerance to the extremely low temperatures (second winter-resistance component). This is evidenced by the high content of the free form of water in the vine of the studied varieties, which characterizes the intensity of metabolic processes in the grape plant and can be associated with hydrothermal conditions in December 2015 (Kushnirenko and Pecherskaya, 1991).

It should be noted the higher content of the bound form of water in the vine of early maturing varieties of Krystall, which suggests that it had a better preparation for the

winter of 2015

In determining the content of pigments (anthocyanins and halcons) - antioxidants that protect cell membranes from destruction (Koshkin, 2010), it was found that the vine grape varieties anthocyanins contain less than halcons. The biggest difference in their content was observed in the variety Krasnostop AZOS and smaller-in the variety Krystall, which may be due to the winter hardiness of varieties.

One of the indicators characterizing frost resistance in the period of organic rest is both the starch content in the vine and its ability to hydrolyze. Determination of the ratio of sucrose to starch in the variety Krystall was 3.8, in the variety Krasnostop AZOS -0.55, which also characterizes the winter hardiness of varieties. A Dostoyney varieties occupies an intermediate position between these varieties (Nenko et al., 2017).

Winter-hardy varieties are characterized by a high content of bound water forms and a great influence on the resistance of plant cells to dehydration have osmoprotectors, such as Proline and sucrose (Jie et al., 2008; Mohammad et al., 2014). Determination of the correlation coefficients between the content of the bound form of water, Proline and sucrose allowed to establish that the more winter-hardy grade Krystall dependence of the bound water on the content of Proline is higher than that of the varieties Dostoyney and Krasnostop AZOS. The accumulation of Proline is accompanied by the prevention of protein denaturation, preservation of the structure and activity of enzymes, which also provides winter hardiness of the variety (Koshkin, 2010; Kuznetsov and Shevyakova, 1999; Barka and Audran, 1997; Ashraf and Foolad, 2007).

These facts suggest a great influence of the genotype of the grape variety, its origin on winter hardiness. Consequently, the presence in the genome of the variety Krystall genetic potential of the Amur grape has a predominant effect on its frost resistance and determines the specificity of metabolic processes. In the pedigree of varieties Dostoyney and Krasnostop AZOS present winter-hardy variety Dzhemete, which determines their winter hardiness. Physiological and biochemical patterns of resistance to low temperatures in these varieties have their own specifics.

One of the elements of resistance of grape varieties to low temperatures is the resistance of the lipid phase of cell membranes to destruction. A large role in preserving the integrity of the lipid phase belongs to the water-soluble antioxidant-ascorbic acid, which is able to restore membrane-bound tocopherol - antioxidant of the lipid phase, which causes a break in the chains of free radical oxidation, interacting with peroxy and alkoxy radicals (Koshkin, 2010; Davey et al., 2000; Foyer et al., 1991).

With artificial freezing of shoots of grapes, the content of bound water in varieties Dostoyney and Krasnostop AZOS increased by 66.7 and 45.5%, respectively and the variety

Krystall did not change, which may be due to the greater stability of the latter. At the same time, hydrolytic processes were activated (the protein and starch content decreased), the Krystall and Dostoyney varieties increased the content of phenolcarboxylic acids, and the grade of Krasnostop AZOS decreased, which may be due to their ecological and geographical origin. Our results are in line with literature. For example, in several studies by Beheshti Rooy et al. (2017) and Wample and Bary (1992), an increase in the concentration of sugar in response to stress caused by low temperature was shown. The authors suggested that sugars can enhance the stabilization of biomembrane by reducing the freezing temperature of intercellular water.

Under cold stress, the content of abscisic acid inducing cold resistance genes increased almost 5-fold in the Krasnostop AZOS variety (Koshkin, 2010). The decrease in the amount of organic acids in the vine of all three varieties after freezing characterizes the decrease in the intensity of breathing.

In February, all grape varieties showed the higher contents of the sum of phenolcarboxylic and ascorbic acids, preventing the damage of cell membranes. The elevated levels of proline contents suggest the enhanced water-retaining capacity of cytoplasm and were used to display their adaptability by the third winter-resistance component. According to some literature data (Yakuba et al., 2015c, d; Foyer, 1993; Foyer et al., 1991), we may assume better resistance of 'Kristall' variety plants to desiccation and formation of proteins with the higher proline contents, strengthening the cell walls in response to stress factors. Accumulation of proline in response to various stress factors, including low-temperature conditions has been correlated with stress tolerance. Proline concentration is higher in stress-tolerant than in stress-sensitive plants (Yakuba et al., 2015c).

In the present study, starch and sucrose content was increased markedly in 'Kristall' variety plants as compared with other varieties. This reflects the higher resistance adaptability of 'Kristall' variety to low temperature stress. Our results are in line with some literature data. For example, Beheshti Rooy et al. (2017) showed the elevation of sugar concentrations in response to stress induced by low temperature. The authors supposed that sugars can increase biomembrane stabilization through decreasing the freezing point of intercellular water.

Our data, together with the literature, indicated the better adaptation of 'Kristall' variety to environmental conditions of Anapa and Taman zone, as compared with 'Krasnostop AZOS' and 'Dostoyney' varieties.

The anatomico-morphological and histochemical studies of the one-year-old vine of the grape varieties under study in March showed them to enter vegetation season, characterized by activity of the growth processes. The starch contents of the fine-cellular core zone in 'Dostoyney', 'Krasnostop AZOS', 'Kristall' varieties was established to be

Table 1: Biochemical indicators of the grape interspecific hybrids adaptation to the stress factors of 2007-2015 winter period.

Biochemical indicators	State of deep dormancy	State of induced dormancy
Water content of shoot,%	13.70– 31.87	16.44 – 34.60
Dry matter contents,%	68.13 – 86.28	65.40– 83.56
Free water contents,%	46.54– 81.25	47.5 – 80.43
Bound water contents,%	18.65 – 53.46	19.85 – 52.5
Ratio of bound water to free water	0.34 – 1.15	0.24 – 1.11
Saccharose contents, mg/g	1.07– 11.83	2.18 – 6.15
Starch contents, mg/g	1.67– 15.24	1.51– 8.51
Proline contents, mg/kg	7.6 – 92.0	46.2 – 190.0
Protein contents, mg/g	2.43– 6.58	2.49 – 8.21
The sum of phenolcarbolic and ascorbic acids, mg/g	0.03 – 13.4	0.01 – 0.06

4.9 points. The anatomo-morphological studies of the buds showed the embryonic processes, responsible for the current year yield. Thus, the obtained results allow supposing the various mechanisms of adaptation of the studied grape varieties to the stress factors of winter period that are typical for Anapa and Taman zone.

The most informative parameters, characterizing the resistance of the studied grape varieties to the stressors of the 2007-2015 winter periods (water content of the shoots, free and bound water contents and their ratio, protein, aminoacids, proline, sugars, the sum of phenolcarbolic and ascorbic acid) are shown in Table 1.

These parameters, indicating physiological and biochemical changes to cold-induced stress could be applied in breeding programs of grape varieties. We assume that the above-mentioned parameters could be useful as markers of tolerance to low-temperature conditions in different climatic regions that are suitable for grape plants growing.

Conclusions

In this study, the key changes of biochemical and morphological characteristics of grape plant of the Anapa and Taman regions were evaluated. The plant response to the different environmental factors showed significant difference among plants of early maturation period (Kristal) as compared with varieties with medium maturation period ('Dostoyiniy' and 'Krasnostop AZOS').

For the interspecific grape hybrids, the optimum physiological and biochemical parameters of winter-resistance (in the state of deep and induced dormancy) and the ranges of their variation, permitting these varieties to survive in extreme conditions of Anapa -Taman zone winter period, were established.

The low temperatures conditions of 2016 brought to insignificant subfreezing of grape plants that may have effect on their production performance. In the state of

induced dormancy under favourable climatic conditions of February 2016, the varieties may be characterized positively by the third winter-resistance component.

Thus, our study indicated significant differences between the studied grape varieties by their ability to adapt to the hydrothermal conditions of the Russian South. The results of the study can be used to monitor frost-resistance of the grape plants for the agricultural and breeding purposes.

REFERENCES

- Arun-Chinnappa KS, Ranawake L, Seneweera S (2017). Impacts and Management of Temperature and Water Stress in Crop Plants. In: Minhas, P., Rane, J., Pasala, R. (eds.) *Abiotic Stress Management for Resilient Agriculture*. Singapore: Springer. pp. 221-233
- Ashraf M, Foolad MR (2007). Roles of glycine betaine and proline in improving plant abiotic stress resistance. *Environ. Exp. Bot.* 59(2):206-216.
- Barka EA, Audran JC (1997). Response of champenoise grapevine to low temperatures: Changes of shoot and bud proline concentrations in response to low temperatures and correlations with freezing tolerance. *J. Horticult. Sci.* 72(4):577-582.
- Beheshti Rooy SS, Hosseini SG, Ghabooli M, Gholami M (2017). Cold-induced physiological and biochemical responses of three grapevine cultivars differing in cold tolerance. *Acta Physiol Plant.* 39(12):264.
- Davey MW, Montagu M, Inze D, Sanmartin M, Kanellis A, Smirnoff N, Benzie IJJ, Strain JJ, Favell D, Fletcher J (2000). Plant L-ascorbic acid: chemistry, function, metabolism, bioavailability and effects of processing. *J. Sci. Food Agric.* 80(7):825-860.
- Dospekhov VA (1979). The methods of a field experiment - M.: Kolos.
- Ferrandino A, Lovisolo C (2014). Abiotic stress effects on grapevine (*Vitis vinifera* L.): Focus on abscisic acid-mediated consequences on secondary metabolism and berry quality. *Environ. Exp. Bot.* 103:138-147
- Foyer CH (1993). Ascorbic acid, in *Antioxidants in Higher Plants*. Ed by Alscher R.G. and Hess J.L., CRC Press, Boca Raton. pp. 32-57.
- Foyer CH, Lelandais M, Edwards EA, Mullineaux P (1991). The role of ascorbate in plants, interactions with photosynthesis, and regulatory significance, in *Active Oxygen. Oxidative Stress and Plant Metabolism*, Ed by Pell E and Steffen K, American Society of Plant Physiologists, Rockville. pp. 131-144.
- Jie Y, Yang H, Zhao H, Zhang W, Li D (2008). Promotion of proline accumulation in apple leaves by bioregulators. *Acta Hort.* 774:237-242.

- Koshkin EI (2010). Physiology of Stability of Agricultural Crops. M.:Drofa.
- Kushnirenko MD, Pecherskaya SN (1991). Physiology of the plants water exchange and drought resistance. Kishinev: "Shtiintsa".
- Kuznetsov VV, Shevyakova NI (1999). Proline under stress: biological role, metabolism, regulation. *Physiol. Plants*. 46(2):321-336.
- Mohammad AH, Anamul H, David JB, Masayuki F (2014). Proline protects plants against abiotic oxidative stress: biochemical and molecular mechanisms: *Oxidative Damage to Plants*. pp. 5:477-522.
- Nenko NI, Ilyina IA, Kiseleva GK, Sudyreva MA (2015). Physiological and biochemical parameters of resistance of grape varieties to the stressors of the winter period in the South of Russia: *Yale J. Sci. Educ. "Yale Univ. Press"*. 1(16):587-598.
- Nenko NI, Petrov VS, Ilyina IA, Kiseleva GK, Sudyreva MA, Sokolova VV (2017). The physiological and biochemical mechanisms of adaptation to the low-temperature stresses of the grape varieties different in ecological and geographical origin. *Horticult. Vinicult.* 5:33-38.
- Ollat N, van Leeuwen C, Garcia de Cortazar-Atauri I, Touzard JM (2017). The challenging issue of climate change for sustainable grape and wine production. *OENO One*. 51:59-60.
- Sha Valli KPS, Nagamallaiiah GV, Dhanunjay RM, Sergeant K, Hausman JF (2014). *Emerging Technologies and Management of Crop Stress Tolerance*. San Diego: Academic Press.
- Swanson CA, El-Shishiny EDH (1958). Translocation of sugars in the Concord grape. *Plant Physiol.* 33:33-37.
- Takahama U, Oniki T (2000). Flavonoides and some other phenolics as substrates of peroxidase: physiological significance of the redox reactions. *Plant Res.* 133(3):301-309.
- The methodological and analytical support of research in horticulture. (2010.) Krasnodar: NCRRIH&V.
- Trejo-Martínez MA, Orozco A, Almaguer-Vargas G, Carvajal-Millán E, Gardea AA, (2009). Metabolic activity of low chilling grapevine buds forced to bud break. *Thermochim. Acta.* 481(1): 28-31.
- Wample RL, Bary A (1992). Harvest Date as a Factor in Carbohydrate Storage and Cold Hardiness of Cabernet Sauvignon Grapevines. *J. Am. Soc. Hort. Sci.* 117(1):32-36.
- Welham SJ, Gezan SA, Clark SJ, Mead A (2014). *Statistical Methods in Biology: Design and Analysis of Experiments and Regression*. New York: Chapman and Hall/CRC.
- Xian M, Luo T, Khan MN, Hu L, Xu Z (2017). Identifying differentially expressed genes associated with tolerance against low temperature stress in brassica napus through transcriptome analysis. *Int. J. Agric. Biol.* 19:273-281.
- Yakuba YF, Ilyina IA, Zakharova MV, Lifar GV (2015). The methodology to determine mass concentration of ascorbic, chlorogenic and caffeic acids in the shoots and leaves of fruit crops and grape-vine with the use of capillary electrophoresis. *Modern instrumental and analytical methods of the fruit crops and grape-vine research*. Krasnodar: NCRRIH&V. pp. 68-72.
- Yakuba YF, Ilyina IA, Zakharova MV, Lifar GV (2015). The methodology to determine mass concentration of cations of ammonium, potassium, sodium, magnesium, calcium in the shoots and leaves of fruit crops and grape-vine with the use of capillary electrophoresis. *Modern instrumental and analytical methods of the fruit crops and grape-vine research*. Krasnodar: NCRRIH&V. pp. 62-67
- Yakuba YF, Ilyina IA, Zakharova MV, Lifar GV (2015). The methodology to determine mass concentration of tartaric, malic, succinic, citric acids in the shoots and leaves of fruit crops and grape-vine with the use of capillary electrophoresis. *Modern instrumental and analytical methods of the fruit crops and grape-vine research*. Krasnodar: NCRRIH&V. pp. 68-79
- Yakuba YF, Ilyina IA, Zakharova MV, Lifar GV (2015). The methodology to determine mass concentration of free amino acids in the shoots and leaves of fruit crops and grape-vine with the use of capillary electrophoresis. *Modern instrumental and analytical methods of the fruit crops and grape-vine research*. Krasnodar: NCRRIH&V. pp. 80-86
- Yegorov EA, Shadrina ZA, Kochyan GA (2017). The model and mechanism of the resource conservation processes management in industrial fruit farming and viticulture. *Academic papers of the NCRRIH&V*. 12:7-12
- Yermakov AI, Arasimovich VE, Smirnova-Ikonnikova MI, Yarosh NP, Lukovnikova GA (1972). *The methods for biochemical study of plants*. Leningrad: Kolos Leningrad Department.
- Zhang J, Wu X, Niu R, Liu Y, Liu N, Xu W, Wang Y (2012). Cold-resistance evaluation in 25 wild grape species. *Vitis*. 51(4):153-160.

Cite this article as:

Nenko NV, Ilyina IA, Kiseleva GK, Yablonskay EK (2019). The low-temperature stress tolerance of the grape varieties of ecological and geographical origin. *Acad. J. Food. Res.* 7(1): 001-011.

Submit your manuscript at

<http://www.academiapublishing.org/journals/ajfr>